

**EXPRESSION OF VASCULAR ENDOTHELIAL GROWTH
FACTOR-A AND ITS RECEPTORS IN UTEROPLACENTAL
TISSUES OF THE NORMOTENSIVE AND HYPERTENSIVE
PREGNANT RAT**

A Thesis Submitted
for the Degree of

MASTER OF SCIENCE

by

HUILING HE

Centre for Bioprocessing and Food Technology

Victoria University of Technology

Melbourne

AUSTRALIA

1999

PREFACE

I hereby certify that this thesis contains no material which has been accepted for the award of any other degree in any university and, that to the best of my knowledge and belief, the thesis contains no material previously published or written by another person, except where due reference is made in the text of this thesis.

The study presented in this thesis was completed by the author at Victoria University of Technology under the supervision of Dr. Kerry Dickson and Dr. Sarah Fraser.

Huiling He
Centre for Bioprocessing and Food Technology,
Victoria University of Technology,
Melbourne, Australia.
1999

ACKNOWLEDGEMENTS

Although my name goes on the cover, this thesis would not be a reality without the hard work and dedication of countless people.

First and foremost, I am very grateful to my principal supervisor, Dr Kerry Dickson, who has offered an opportunity for me to study at Victoria University and provided financial support during my second year of study. Many thanks for her support, guidance, encouragement, constructive criticism throughout the course of my study, and critical reading of the manuscript.

I am sincerely indebted to my co-supervisor, Dr Sarah Fraser, who has provided invaluable supervision through her wealth of knowledge in molecular biology and development of new techniques. She has made a major contribution to the success of this thesis. The time that we have shared in the laboratory including evenings and weekends will be forever imprinted in my memory as some of the best of my professional career.

I would like to express my thanks to Professor Bob Fairclough, for the invaluable encouragement and emotional support, enabling me to complete my study.

I would like to convey my thanks to Professor Margaret Britz, the former Director of the Centre for Bioprocessing and Food Technology (CBFT), for supporting this work.

I would like to thank Dr David Finkelstein, Department of Medicine, Monash University, for allowing me to use his laboratory and equipment to collect the rat tissues. This project would not have proceeded without his help.

I would like to thank Professor Graham Jenkin, Department of Physiology, Monash University, for the initial stimulus to work on VEGF-A, and for providing me with the initial VEGF-A primers.

I would like to thank Ms Usula Manuepillai for kindly providing the human G3PDH primers and for sequencing the resultant RT-PCR product.

I would like to acknowledge Dr Mrinal Bhave, for her guidance and help during the initial stages of this project, and for her kind donation of a clone G3PDH plasmid.

I would like to thank Dr Tseng Lau, who has kindly provided assistance for the immunohistochemistry and *in situ* hybridisation experiments.

I am particularly grateful to my best friend, Helen Chunying Feng, for her enthusiastic help and technical advise, particularly for her assistance in the initial RNA work, and for her mental stimulus that has led me to complete this study.

My sincere thanks go to Ms Hong Wang from the Institute for Reproduction and Development, Monash University, who has provided enthusiastic assistance and excellent technical advice in immunohistochemistry and *in situ* hybridiazation.

Many thanks to the lovely friends and colleagues at the CBFT and the School of Life Sciences. I would like to thank Karoline Tellbach, for her help with the administration of anaesthesia, the holding of the rats, the preparation of surgical equipment; and for her warm help and sharing in the laboratory.

I would like to thank Matthew Knight, for his warm advise and wonderful friendship. I would also like to thank Hemalatha Jegasothy; Aslam Khan and Diana Emslie for their kindness and for sincerely caring.

Finally, I would like to dedicate this thesis to my husband John, for his understanding and full support especially as we experienced extreme financial difficulty and the project took away all my attention and time. Even though he suffered from severe illness in hospital, he still encouraged me to concentrate on my study. I also want to dedicate this project to my family members, especially my Dad, from him came all care and blessings.

CONFERENCE PRESENTATIONS

Part of this work has been presented at the following conferences

H. L. He, S. M. Fraser, K. A. Dickson. mRNA Expression of Vascular Endothelial Growth Factor (VEGF) and its Receptor (Flt-1) in Uteroplacental Tissues of the Pregnant Spontaneously Hypertensive Rat (SHR). *Fetal and Neonatal Physiological Society 25th Annual Meeting*. California, U.S.A, pp 42, 1998.

H. L. He, S. M. Fraser, K. A. Dickson. mRNA Expression of Vascular Endothelial Growth Factor (VEGF) and its Receptor (Flt-1) in Uteroplacental Tissues of the Pregnant Spontaneously Hypertensive Rat (SHR). *Fertility Society Association Conference*. Hobart, Australia, pp 72, 1998.

ABBREVIATIONS

aa	Amino acid
A₂	Thromboxane
H₂O₂	Hydrogen peroxide
ANGIS	Australian National Genomic Information Service
Ang-1	Angiopoietin 1
Ang-2	Angiopoietin 2
β-FGF	β fibroblast growth factor
bFGF	Basic fibroblast growth factor
BSA	Bovine serum albumin
cDNA	Complementary deoxyribonucleic acid
CT	Cytotrophoblast
DDT	Dithiothreitol
DEPC	Diethylpyrocarbonate
DIG	Digoxigenin
DNA	Deoxyribonucleic acid
DNase	Deoxyribonuclease
dNTPs	Equimolar mixture of dATP, dCTP, dGTP, dTTP
EB	Ethidium bromide
ECM	Extracellular matrix
EDTA	Ethylenediaminetetra-acetic acid
EGF	Epidermal growth factor
EP	Epithelial cell

Flk-1	Fetal liver kinase (rat)
Flt-1	c-fms-like tyrosine kinase
G3PDH	Glyceraldehyde-3-phosphate dehydrogenase
HCG	Human chorionic gonadotrophin
HGF	Hepatocyte growth factor
IFNα	Interferon alpha
IL	Interleukin
IUGR	Intrauterine growth retardation
KDR	Kinase insert domain-containing receptor (human)
MMP	Metalloproteinase
mRNA	Messenger ribonucleic acid
NO	Nitric oxide
PA	Plasminogen activator inhibitors
PAI	Plasminogen activator inhibitors
PBS	Phosphate buffered saline
PDGF	Platelet derived endothelial cell growth factor
PGI₂	Prostacyclin
PIGF	Placental growth factor
RNase	Ribonuclease
RTK	Receptor tyrosine kinase
RT-PCR	Reverse transcriptase polymerase chain reaction
SHR	Spontaneously hypertensive rat
TGF-α	Transforming growth factor alpha
TIMP	Tissue inhibitor of metalloproteinase

TNFα	Tumour necrosis factor alpha
TSP1	Thrombospondin-1
VEGF	Vascular endothelial growth factor
VEGFR-1	Vascular endothelial growth factor receptor 1
VEGFR-2	Vascular endothelial growth factor receptor 2
VEGF-A	Vascular endothelial growth factor A
VEGF-B	Vascular endothelial growth factor B
VEGF-C	Vascular endothelial growth factor C
VEGF-D	Vascular endothelial growth factor D
VPF	Vascular permeability factor
UV	Ultraviolet
WKY	Wistar Kyoto rat

TABLE OF CONTENTS

	Page
PREFACE	ii
ACKNOWLEDGEMENTS	iii
CONFERENCE PRESENTATIONS	v
ABBREVIATIONS	vi
TABLE OF CONTENTS	ix
LIST OF TABLES	xvi
LIST OF FIGURES	xix
SUMMARY	1
CHAPTER 1: LITERATURE REVIEW	3
1.0.0 INTRODUCTION	4
1.1.0 NORMAL PLACENTAL DEVELOPMENT	5
1.1.1 Structure of the Uteroplacental Circulation	5
1.1.2 Uteroplacental Blood Flow	6
1.2.0 IMPAIRED PLACENTAL DEVELOPMENT WITH PRE-ECLAMPSIA	6
1.2.1 Structure of the Uteroplacental Circulation with Pre-eclampsia	9
1.2.2 Uteroplacental Blood Flow with Pre-eclampsia	9
1.3.0 ANIMAL MODEL FOR HYPERTENSION DURING PREGNANCY	10
1.4.0 ANGIOGENESIS	12
1.4.1 Mechanisms of Angiogenesis	13
1.4.2 Role of Angiogenic and Anti-Angiogenic Factors	15
1.4.3 Role of Physical Stress and Blood Flow	17

1.4.4	Role of Monocytes	17
1.4.5	Role of Hormones	17
1.4.6	Role of Hypoxia	17
1.5.0	VASCULAR ENDOTHELIAL GROWTH FACTOR (VEGF) FAMILY	18
1.5.1	Vascular Endothelial Growth Factor A (VEGF-A)	18
1.5.2	Vascular Endothelial Growth Factor B (VEGF-B)	19
1.5.3	Vascular Endothelial Growth Factor C (VEGF-C)	19
1.5.4	Vascular Endothelial Growth Factor D (VEGF-D)	20
1.5.5	Placental Growth Factor (PIGF)	20
1.6.0	STRUCTURE OF VEGF-A	21
1.7.0	VEGFR-1 AND VEGFR-2	26
1.8.0	FUNCTIONS OF VEGF-A	34
1.8.1	Endothelial Cell Growth and Proliferation	34
1.8.2	Breakdown of the Endothelial Basement Membrane	35
1.8.3	Increasing Vascular Permeability and Protein Leakage	35
1.8.4	Monocyte Chemotaxis	36
1.8.5	Changes in Vascular Tone	36
1.9.0	REGULATION OF VEGF-A	37
1.9.1	Growth Factors	37
1.9.2	Physical Stress and Blood Flow	39
1.9.3	Hormones	39
1.9.4	Hypoxia	39
1.9.5	Erythropoietin	40
1.9.6	Cell Differentiation	40
1.10.0	VEGF-A IN UTEROPLACENTAL TISSUES	40

1.11.0	AIMS	42
1.11.1	Specific Aims	43
CHAPTER 2: MATERIALS AND METHODS		44
2.1.0	ANIMALS	45
2.1.1	Animal Husbandry	45
2.1.2	Ethical Approval	45
2.1.3	Collection of Uteroplacental Tissues	45
2.2.0	RNA EXTRACTION	48
2.2.1	Total RNA Extraction by the Guanidinium Technique	49
2.2.2	Total RNA Extraction by the Trizol Technique	49
2.2.3	mRNA Extraction by the Poly-AT Tract Isolation Technique	50
2.3.0	QUALITY AND QUANTIFICATION OF RNA	51
2.3.1	Quality of RNA by Electrophoresis	51
2.3.1.1	Denaturing gel electrophoresis	51
2.3.1.2	Non-denaturing gel electrophoresis	51
2.3.2	Concentration and Purity of RNA	52
2.4.0	REVERSE TRANSCRIPTASE POLYMERASE CHAIN REACTION	52
2.4.1	Design of Primers	52
2.4.1.1	G3PDH primers	52
2.4.1.2	VEGF-A primers	53
2.4.1.3	VEGFR-1 primers	54
2.4.1.4	VEGFR-2 Primers	54
2.4.2	DNase Treatment of RNA	55
2.4.3	First Strand cDNA Synthesis Using SUPERSCRIPT™ II RT	55
2.4.4	PCR Amplification	56
2.4.4.1	G3PDH	57
2.4.4.2	VEGF-A (primer <i>set 1</i>)	58

2.4.4.3	VEGF-A (primer <i>set 2</i>)	59
2.4.4.4	VEGFR-1	60
2.4.4.5	VEGFR-2	60
2.5.0	SEQUENCE ANALYSIS	62
2.6.0	DENSITOMETRY	62
2.7.0	STATISTICAL ANALYSIS	63
2.8.0	PROBES FOR NORTHERN BLOT ANALYSIS	63
2.8.1	G3PDH	63
2.8.2	VEGF-A	63
2.8.3	VEGFR-1	65
2.8.4	VEGFR-2	67
2.9.0	DNA PURIFICATION	67
2.10.0	CLONING OF PROBES	67
2.10.1	Ligation	68
2.10.2	Transformation	68
2.10.3	Alkaline Lysis Miniprep	68
2.11.0	NORTHERN BLOTTING	69
2.11.1	Preparation of Blots	69
2.11.2	Radioactive Labelling of cDNA Probes	70
2.11.3	Non-radioactive Labelling of cDNA Probes	70
2.11.4	Northern Hybridisation Using Radioactive cDNA	72
2.11.4.1	pre-hybridisation	72
2.11.4.2	hybridisation	72
2.11.4.3	post-hybridisation	73
2.11.4.4	autoradiography	73

2.11.5.0	Northern Hybridisation Using Non-Radioactive cDNA Probes	73
2.11.5.1	pre-hybridisation	73
2.11.5.2	hybridisation	74
2.11.5.3	post-hybridisation	74
2.11.5.4	immunological and chemiluminescence detection	74
2.12.0	IMMUNOHISTOCHEMISTRY	75
2.12.1	Frozen Tissues	75
2.12.2	Fixed Tissues	75
2.12.3	VEGF-A Immunohistochemistry	75
2.13.0	<i>IN SITU</i> HYBRIDIZATION	77
2.13.1	Preparation of Riboprobe	77
2.13.2	Non-radioactive Labelling of Riboprobes for <i>In situ</i> Hybridisation	78
2.13.3	Northern Blot Analysis using Riboprobes	78
2.13.4	<i>In situ</i> Hybridisation	79
	CHAPTER 3: ESTABLISHMENT OF RT-PCR, NORTHERN BLOT ANALYSIS, IMMUNOHISTOCHEMISTRY AND IN SITU HYBRIDISATION TECHNIQUES TO MEASURE EXPRESSION AND LOCALISATION OF VEGF-A, VEGFR-1 AND VEGFR-2 IN NORMAL UTEROPLACENTAL RAT TISSUES AT EARLY, MID AND LATE GESTATION.	80
3.1.0	INTRODUCTION	81
3.2.0	METHODS	82
3.3.0	RESULTS	82
3.3.1	Quality and Quantification of RNA Extraction	82
3.3.2	Reverse Transcriptase Polymerase Chain Reaction	85

3.3.2.1	DNase treatment	85
3.3.2.2	G3PDH RT-PCR	85
3.3.2.3	VEGF-A RT-PCR	88
3.3.2.4	VEGFR-1 RT-PCR	88
3.3.2.5	VEGFR-2 RT-PCR	98
3.3.3	VEGF-A and VEGFR-1 Expression, Using RT-PCR, at Early, Mid and Late Gestation	98
3.3.4	Semi-quantitative Analysis of VEGF-A and VEGFR-1 Expression	102
3.3.5	Northern Blot Analysis	105
3.3.5.1	VEGF-A	105
3.3.5.2	VEGFR-1	105
3.3.6	VEGF-A Immunohistochemistry	106
3.3.7	VEGF-A <i>In situ</i> Hybridisation	106
3.4.0	DISCUSSION	117
3.4.1	Extraction of Total and mRNA	118
3.4.2	RT-PCR	118
3.4.3	Northern Blot Analysis	121
3.4.4	Immunohistochemistry	122
	CHAPTER 4: EXPRESSION OF VEGF-A AND VEGFR-1 IN UTEROPLACENTAL TISSUES FROM NORMOTENSIVE (WKY) AND SPONTANEOUSLY HYPERTENSIVE (SHR) RATS	123
4.1.0	INTRODUCTION	124
4.2.0	METHODS	124
4.3.0	RESULTS	124
4.3.1	Animals	124
4.3.2	VEGF-A RT-PCR	124
4.3.3	VEGF-A Northern Blot Analysis	129

4.3.4	VEGFR-1 RT-PCR	129
4.3.5	VEGFR-2 RT-PCR	129
4.4.0	DISCUSSION	135
4.4.1	SHR as a Model for Pre-eclampsia	135
4.4.2	VEGF-A, VEGFR-1 and VEGFR-2 in SHR and WKY Rats	136
4.4.3	Relationship between VEGF-A and Pre-eclampsia	139
CHAPTER 5: CONCLUSIONS AND FURTHER STUDIES		142
REFERENCES		144
SOLUTIONS AND BUFFERS		169

LIST OF TABLES

	Page
1.1. Summary of the main factors involved in capillary sprouting.	16
1.2. Summary of various properties of the VEGF family members.	18
1.3. Size of VEGF-A mRNA transcripts from the literature.	22
1.4. Size of the VEGFR-1 mRNA transcripts from the literature.	27
1.5. Size of VEGFR-2 mRNA transcripts from the literature.	27
2.1. Number of normal Sprague Dawley rats obtained at each gestational age.	48
2.2. Number of normotensive (WKY) and spontaneously hypertensive (SHR) rats obtained at each gestational age.	48
2.3. Theoretical sizes of RT-PCR products using <i>set 1</i> and <i>set 2</i> VEGF-A primers.	54
2.4. First strand cDNA synthesis using SUPERSCRIPT™ II RT	56
2.5. The composition of the PCR reaction for the amplification of G3PDH.	57
2.6. PCR cycling parameters for G3PDH	57
2.7. The composition of the PCR reaction for the amplification of VEGF-A (primer <i>set 1</i>).	58

2.8.	PCR cycling parameters for VEGF-A (primer <i>set 1</i>).	58
2.9.	The composition of the PCR reaction for the amplification of VEGF-A (primer <i>set 2</i>).	59
2.10.	PCR Cycling parameters for VEGF-A (primer <i>set 2</i>).	59
2.11.	The composition of the PCR reaction for the amplification of VEGFR-1.	60
2.12.	PCR cycling parameters for VEGFR-1.	60
2.13.	The composition of the PCR reaction for the amplification of VEGFR-2.	61
2.14.	PCR cycling parameters for VEGFR-2.	61
2.15.	PCR cycling parameters for sequencing.	62
2.16.	The composition of the PCR reaction for the amplification of the VEGF-A probe.	64
2.17.	PCR cycling parameters for the VEGF-A probe.	64
2.18.	DIG quantification test strip procedure.	71
2.19.	Post-hybridisation washing protocol for radioactive cDNA probes.	73
2.20.	Post-hybridisation washing protocol for non-radioactive cDNA probes.	74
2.21.	The composition of the PCR reaction for amplification of the VEGF-	77

A riboprobe.

2.22.	PCR cycling parameters for the VEGF-A riboprobe.	78
3.1.	Comparison of yield and purity of total RNA extractions using the Guanidinium thiocyanate and Trizol techniques.	82
3.2.	Ratio of VEGF-A ₁₆₄ to G3PDH in uteroplacental tissues at 5, 11 and 21 days of gestation assessed by laser densitometry of RT-PCR products.	100
4.1.	Litter size for WKY and SHR rats at 7, 11 and 19 days of gestation.	124
4.2.	Ratio of VEGF-A to G3PDH for one WKY and one SHR rat at early, mid and late gestation.	126

LIST OF FIGURES

	Page
1.1. Relationship between the fetal and maternal portions of the full-term placenta.	7
1.2. Structure of a chorionic villus.	8
1.3. VEGF-A splice variants in the human.	23
1.4. The sequence of human VEGF-A ₂₀₆ cDNA and comparison of amino acid sequence of multiple VEGF-A molecular species.	24
1.5. DNA sequence and predicted translation products of murine VEGF-A.	25
1.6. Structure of receptor tyrosine kinases.	29
1.7. Structure of VEGFR-1 (or Flt-1) and VEGFR-2 (or KDR).	30
1.8. Partial VEGFR-1 amino acid sequence (286) from human placentae.	31
1.9. VEGFR-2 amino acid sequence from mouse.	32
1.10. Interaction of VEGF-A with its receptors VEGFR-1 and VEGFR-2.	33
1.11. Schematic model for the complex series of effects of VEGF-A on the vascular endothelium.	38
2.1. Collection of uteroplacental tissues.	46

2.2.	Schematic diagram showing the technique for freezing tissues in OCT.	47
2.3.	Gels showing purified VEGF-A ₁₆₄ DNA fragments.	66
2.4.	Photograph showing DIG quantification test strip.	72
3.1.	Electrophoresis gels of total RNA extraction using the Guanidinium thiocyanate and Trizol techniques.	83
3.2.	Electrophoresis gel of mRNA extracted from total RNA.	84
3.3.	Dependence of PCR product formation on the presence of reverse transcriptase.	86
3.4.	RT-PCR analysis of G3PDH expression in uteroplacental tissues.	87
3.5.	RT-PCR analysis of VEGF-A expression in placental and uterine tissues at 19 days of gestation.	89
3.6.	Partial sequence alignment of the VEGF-A ₁₈₈ (308 bp) RT-PCR product (primer <i>set 1</i>).	90
3.7.	Partial sequence alignment of the VEGF-A ₁₆₄ (236 bp) RT-PCR product (primer <i>set 1</i>).	91
3.8.	RT-PCR analysis of VEGF-A expression (primers <i>set 2</i>) in uteroplacental tissues.	92
3.9.	Partial sequence alignment of the VEGF-A ₁₂₀ (364 bp) RT-PCR product (primer <i>set 2</i>).	93

3.10.	RT-PCR analysis of VEGFR-1 expression in uterine tissues at 19 days of gestation.	94
3.11.	Partial sequence alignment of the VEGFR-1 (280 bp) RT-PCR product.	95
3.12.	RT-PCR analysis of VEGFR-2 expression in uteroplacental tissues.	96
3.13.	Partial sequence alignment of the VEGFR-2 (383 bp) RT-PCR product.	97
3.14.	RT-PCR analysis of VEGF-A and G3PDH expression in uteroplacental tissues at day 5, 11 and 21 days of gestation.	99
3.15.	RT-PCR analysis of VEGFR-1 and G3PDH expression in uteroplacental tissues at day 5, 11 and 21 days of gestation.	101
3.16.	Kinetics of RT-PCR for VEGF-A and G3PDH.	103
3.17.	Kinetics of RT-PCR for VEGFR-1 and G3PDH.	104
3.18.	Northern blot analysis of VEGF-A and G3PDH expression in uteroplacental tissues over a range of gestational ages.	107
3.19.	Northern blot analysis of VEGF-A and G3PDH expression in uteroplacental tissues over a range of gestational ages.	108
3.20.	Northern blot analysis of VEGF-A and G3PDH expression in uterine tissues from day 13 to 21 of gestation.	109
3.21.	Northern blot analysis of VEGF-A and G3PDH expression in placental tissues from day 13 to 21 of gestation.	110

3.22.	Northern blot analysis of VEGF-A expression in uterine and placental tissues at day 19 of gestation.	111
3.23.	Northern blot analysis of VEGF-A expression in uterine tissues.	112
3.24.	Northern blot analysis of VEGFR-1 expression in uteroplacental tissues at day 7, 11 and 19 days of gestation.	113
3.25.	VEGF-A protein localisation in maternal decidua as revealed by immunohistochemistry.	114
3.26.	VEGF-A protein localisation in maternal decidua as revealed by immunohistochemistry.	115
3.27.	Negative control for immunohistochemistry.	116
4.1.	Litter size for WKY and SHR rats at day 7, 11 and 19 of gestation.	125
4.2.	RT-PCR analysis of VEGF-A (primer <i>set 1</i>) and G3PDH expression in uteroplacental tissues from WKY and SHR rats at 7, 11 and 19 days of gestation.	127
4.3.	RT-PCR analysis of VEGF-A (primer <i>set 2</i>) and G3PDH expression in uteroplacental tissues from WKY and SHR rats at 11 and 19 days of gestation.	128
4.4.	Northern blot analysis of mRNA expression of VEGF-A and G3PDH in WKY and SHR rats at 7, 11 and 19 days of gestation.	130
4.5.	Northern blot analysis of mRNA expression of VEGF-A and G3PDH in a second WKY and SHR rat at 7, 11 and 19 days of gestation.	131

- 4.6. RT-PCR analysis of VEGFR-1 and G3PDH, in uteroplacental tissues 132
from WKY and SHR rats at 7, 11 and 19 days of gestation.

- 4.7. RT-PCR analysis of VEGFR-1 and G3PDH in uteroplacental tissues, 133
for a second WKY and SHR rat at 7, 11 and 19 days of gestation.

- 4.8. RT-PCR analysis of VEGFR-2 and G3PDH in uteroplacental tissues, 134
from WKY and SHR rats at 7, 11 and 19 days of gestation.

SUMMARY

Hypertension, is a common clinical complication of pregnancy, often leading to pre-eclampsia and fetal intrauterine growth retardation. Many studies have shown that vascular endothelial growth factor A (VEGF-A) is a potent angiogenic factor, that is, it is responsible for the formation of new blood vessels from existing vessels. VEGF-A has been shown to be expressed in uteroplacental tissues, particularly during implantation. Levels of VEGF-A mRNA have also been shown to be significantly lower in placental tissue from pre-eclamptic women compared with control women (Cooper *et al*, 1996). The first aim of this project, was to establish the use of RT-PCR (reverse transcriptase-polymerase chain reaction), Northern blot analysis, immunohistochemistry and *in situ* hybridization, to measure the expression and localization of VEGF-A, vascular endothelial growth factor receptor 1 (VEGFR-1) and vascular endothelial growth factor receptor 2 (VEGFR-2) in normal rat uteroplacental tissues at early, mid and late gestation. The second aim of this project was to measure expression of VEGF-A, VEGFR-1 and VEGFR-2 in uteroplacental tissues of the normotensive (WKY) and the spontaneously hypertensive rat (SHR) at 7, 11 and 19 days of gestation. Spontaneously hypertensive rats have been previously established as an animal model for pre-eclampsia.

The uterus containing the placenta and fetus was removed from normal Sprague Dawley, WKY and SHR rats over a range of gestational ages. After 11 days of gestation, the uterus and placenta were separated. Tissues were frozen immediately in liquid nitrogen and stored at -80°C , or fixed in 10% formaldehyde for immunohistochemistry. Total RNA was extracted using TRIZOL reagent and mRNA levels of VEGF-A and its receptors were examined using RT-PCR and Northern blot analysis. mRNA levels of VEGF-A and its receptors were compared with those of the housekeeping gene glyceraldehyde 3-phosphate dehydrogenase (G3PDH).

We have successfully used RT-PCR to measure mRNA expression of VEGF-A, VEGFR-1 and VEGFR-2; Northern blot analysis to measure expression of VEGF-A, and immunohistochemistry to localize VEGF-A protein in rat uteroplacental tissues. We have shown that VEGF-A and its receptors were expressed in uteroplacental

tissues of the normal rat. We found no major differences in the expression levels of VEGF-A and VEGFR-1 in uteroplacental tissues at early, mid and late gestation. We showed that VEGF-A₁₆₄ was the predominant isoform found in uteroplacental tissues. Finally, we found no major differences in the expression levels of VEGF-A, VEGFR-1 and VEGFR-2 in uteroplacental tissues in normotensive rats (WKY) compared with hypertensive rats (SHR).

We conclude that VEGF-A and its receptors, VEGFR-1 and VEGFR-2, were expressed in uteroplacental tissues of pregnant rats. Furthermore, mRNA levels of VEGF-A and its receptors were comparable in normotensive and hypertensive rats. These results suggest that VEGF-A and its receptors may have an important role in the development of the fetal and maternal portions of the placenta, but that they are not the primary factors involved in the aetiology of pre-eclampsia.